# Neurochemistry of Nigral Degeneration

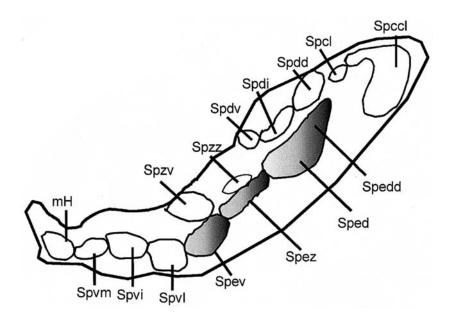
#### Jayaraman Rao

Louisiana State University Health Sciences Center, New Orleans, Louisiana, U.S.A.

# INTRODUCTION

Progressive degeneration of the dopaminergic neurons of the substantia nigra is the pathological hallmark of idiopathic Parkinson's disease (PD). Based on cytoarchitectonics and melanization, Hassler divided the human substantia nigra into many subsections and demonstrated that the ventral and lateral regions of the substantia nigra may be preferentially involved in early stages of the disease (1,2) (Fig. 1), an observation that was later confirmed in human PD (3,4). The etiology of the progressive degeneration of the substantia nigra pars compacta cells is unknown. It has been proposed that the clinical signs and symptoms of PD emerge only after a loss of 75% of the nigral neurons. Positron emission tomography (PET) and single photon emission computed tomography (SPECT) studies also indicate that the rate of loss of dopaminergic neurons is about 6-13% in patients with PD compared to 0-2.5% in healthy controls (5). These facts suggest that the process of degeneration of nigral neurons is initiated several years ahead of the onset of the clinical expression of the disease.

Epidemiological evidence emphasizes the role of environmental toxins in the development of PD. Discoveries of gene mutations responsible for



**FIGURE 1** A diagrammatic representation of the subdivisions of human substantia nigra as defined by Hassler. The shaded subnuclei in the ventral and lateral regions degenerate selectively in the early stages of PD. (Adapted from Refs. 1, 2.)

inherited forms of PD have increased interest in the role of genetics in the etiology of PD. Modern molecular biological approaches are pointing to the possibility that dysfunction of a variety of cellular mechanisms may result in an insidious and a slowly progressive levodopa-responsive parkinsonism that is indistinguishable from PD. These observations raise the issue of whether there may be multiple etiologies for PD. In this chapter, some of the neurochemical changes noted in the degenerating dopaminergic neurons of the substantia nigra in experimental models of PD, inherited forms of PD, and idiopathic PD will be summarized.

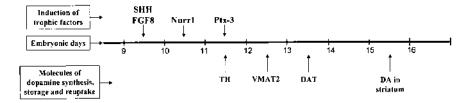
#### **GROWTH FACTORS AND THE SUBSTANTIA NIGRA**

Neurons and glia need a very small concentration of trophic or growth factor(s) for their continued existence, maintenance of a normal connectivity and physiological state, as well as recovery from chemical and physical injury (6). If these growth factors are withdrawn or if their continuing influence is affected both neurons and glia may fail to differentiate and die during embryogenesis. As adult neurons, they may lose their efficiency or

become vulnerable to toxic factors and ultimately die. Under stressful circumstances, neurons and glia upregulate the expression of growth factors and their specific receptors or acquire them from their target neurons and the surrounding glial cells and recover from the injury. Age-dependent decline and withdrawal of neurotrophins and their receptors (7,8) are thought to play an important role in the pathogenesis of Alzheimer's disease. The dopaminergic neurons of the substantia nigra are dependant on several such trophic and growth factors for normal ontogenesis and survival as fully differentiated mature adult neurons. In experimental models of PD and in idiopathic PD, the dopamine neurons of the substantia nigra show a deficiency of expression of several growth factors. It is possible that withdrawal of these growth factors in the dopaminergic neurons of the substantia nigra may contribute to the pathogenesis of idiopathic PD.

# Trophic Factors and Ontogenesis of Dopaminergic Neurons of the Substantia Nigra

The normal development of dopaminergic neurons in ventral mesencephalon depends on the influence of many trophic factors. Between embryonic days E9.5 and E16, several trophic factors play important roles in the induction, differentiation, as well as complete maturation of dopamine synthesis, release, and reuptake machinery of ventral mesencephalic dopamine neurons (Fig. 2). Around the embryonic day of E 9.5, two such factors, namely sonic hedgehog (SHH) (9) and fibroblast growth factor 8 (FGF8), define the site of induction of dopamine neurons in the ventral mesencephalon. SHH is a member of the hedgehog family of signaling protein that plays a major role in the differentiation of diverse groups of neurons in the ventral half of the neural tube, including dopamine and serotoninergic neurons (10). SHH may even play an important role in maintenance of adult dopamine neurons since intrastriatal injection of SHH diminishes the motor behavioral defects of 6-hydroxydopamine (6-OHDA) models of PD (11).



**FIGURE 2** The ontogenesis of dopaminergic neurons of substantia nigra in rats and mice. (Adapted from Refs. 9, 12, 26, 27, 28.)

Around embryonic day E10.5, Nurrl is expressed in a group of cells in the ventral mesencephalic region (12). Nurrl is a member of the "orphan receptors" transcription factors and is expressed in several areas of the brain, but it is expressed intensely and selectively in the dopaminergic neurons of the ventral mesencephalon (12,13). Nurrl knockout results in the complete lack of development of dopaminergic neurons in the midbrain (14,15) with almost 98% reduction of dopamine in the striatum (16). Nurrl and tyrosine hydroxylase (TH) are coexpressed, with the maximum intensity in the midbrain substantia nigra neurons (17-19). The expression of dopamine transporter molecule may also be dependant on the Nurrl gene (20). Nurrl is expressed even in adult neurons, suggesting a role for a continuing influence on TH expression and survival of adult nigral dopamine neurons (21). Nurrl-deficient animals are more susceptible to 1methyl-4-phenyl-1,2,3,6-tetrahydropyridine (MPTP) treatment (22), and Nurrl polymorphism may be associated with familial and idiopathic PD (23).

One day after expression of Nurrl, on E11.5, another factor, Pentraxin 3 (Ptx3), is expressed solely in the dopaminergic neurons of the ventral tegmental area and the substantia nigra (24). Ptx3 is reduced in the 6-OHDA model of PD as well as in idiopathic PD (24). The expression of Ptx3 in these neurons coincides with the expression of the TH gene in the same neurons (25). The appearance of Nurrl and Ptx3 early in the ontogenesis of dopaminergic neurons suggests that these factors are responsible for differentiation and maturation of ventral mesencephalic dopamine neurons rather than establishment of connectivity to the striatum. The expression and maturation of other components of dopamine release, reuptake, and storage are completed by the embryonic day E16, and the release of dopamine in the striatum is first noted around E17 (26–28).

Besides SHH, Nurr-1, and Ptx-3, NFIAb (29), LMX1b (30), heparinbinding epidermal growth factor-like growth factor (HB-EGF) (31), En-1, and En-2 of the engrailed gene family (32) have roles in the survival of the midbrain dopaminergic system during ontogenesis. En-1 and En-2 also appear to regulate the expression of  $\alpha$ -synuclein, the major constituent of Lewy bodies in PD (32).

# **Growth Factors and Nigral Injury**

Besides these trophic factors that play important roles in the ontogenesis of nigral dopamine neurons as well as maintaining the survival of differentiated adult dopamine neurons in the nigra, several other neurotrophic factors may play a neuroprotective role in many models of injury to the dopaminergic neurons of the substantia nigra (8,33). These neurotrophic factors have significant structural and functional similarities and are classified into the nerve-growth factor (NGF) superfamily, the glial-derived neurotrophic factor (GDNF) family, the neurokine family, and the nonneuronal growth factors.

The superfamily of neurotrophins includes the NGF, brain-derived neurotrophic factor (BDNF), and neurotrophins (NT) 3, 4/5, and 6. All members of the neurotrophin family interact with two structurally unrelated receptors, namely the tyrosine kinases (trkA, trkB, and trkC) and the lowaffinity binding neurotrophin receptor p75<sup>NTR</sup> (34,35). Recent studies have shown that the *trk* and  $p75^{NTR}$  are coexpressed in the same cell and that the molecular signaling pathways of these two receptors may be stimulated independently or coactivated (36,37). Even though stimulation of these two pathways may interact with each other, they may have opposite effects. Stimulation of the *trk* receptors subserves a neuroprotective effect by inactivating several factors that are apoptotic (35). The p75<sup>NTR</sup> is structurally unlike the *trk* receptors. It belongs to the tumor necrosis factor (TNF) receptor family and contains the "death domain" (35,38). Stimulation of p75<sup>NTR</sup> has been shown to induce several molecules that can initiate apoptosis, but under other conditions p75<sup>NTR</sup> stimulation can be antiapoptotic (34,35).

The dopaminergic neurons of the substantia nigra in humans immunostain for proteins of FGF, BDNF, and NT3 as well as trkA, trkB, and trkC. The immunoreactivity is noted more intensely in the medial areas of the nigra than the lateral areas (39), providing one piece of evidence for selective loss of neurons in the lateral regions. BDNF, when injected into the striatum, is transported only to the soma of dopaminergic neurons of the substantia nigra. These findings suggest that BDNF found in nigral neurons may be synthesized locally within the neurons or can be acquired by retrograde transport from the striatal neurons (40). The melanized dopamine-containing neurons of the substantia nigra show a greater loss of BDNF than the nonmelanized cells (41,42). BDNF prevents nigral degeneration induced in MPTP models of PD (43).

GDNF, a neurotrophic factor belonging to the TGF- $\beta$  superfamily, and its receptors, Ret and GDNF- $\alpha$ 1, are expressed highly in the dopaminergic neurons of the substantia nigra (8,33). GDNF concentrations are decreased in the nigral neurons in PD (44). When injected in the striatum, GDNF is selectively transported retrogradely to nigral dopaminergic neurons (40). GDNF, like BDNF, protects mesencephalic dopamine neurons from 6-OHDA and MPTP toxicity and improves motor functions in these models of PD (45,46). However, the molecular pathways that mediate neuroprotective effects of these two neurotrophic factors may be different (47). Neurokines are neuropoietic cytokines. Among the neurokines, an increased level of interleukin-6 (IL-6) has been demonstrated in the striatum. Many cytokines, which are traditionally recognized to play antiinflammatory roles outside the brain, have now been recognized to be expressed in the brain (48) in response to tissue injury or inflammation (e.g., multiple sclerosis) (49). Some of these cytokines may be neuroprotective and others apoptotic. In this regard, levels of TNF- $\alpha$ , TGF- $\alpha$ , TGF- $\beta$ , IL-1 $\beta$ , IL-4, and IL-6 are elevated in the striatum in PD. This has led to the hypothesis that the pathogenesis of PD may be the result of an imbalance between the actions of the antiapoptotic neurotrophic factors and the proapoptotic factors (50,51).

Several neurotrophic factors have been recognized to be expressed by nonneural cells. Among these, basic fibroblast growth factor (bFGF or FGF2) has been demonstrated to be expressed in the substantia nigra (52), and a profound depletion of bFGF was noted in the nigral neurons in PD (53,54). Acute and intermittent injections of nicotine increase FGF2 expression and mediate neuroprotective effects in several models of neuronal injury (55,56) including 6-OHDA and MPTP models of PD (57–59).

# Summary

It is important to point out that it is unclear whether the reduction of the neurotrophic factors noted in the nigral neurons of PD is the cause or a consequence of nigral degeneration. While in animal models of acute injury the striatum and the dopaminergic neurons of the substantia nigra may be protected by these neurotrophic factors, the role played by these factors in PD remains to be established. Even if the observed decrease in neurotrophic factors and their receptors is a consequence of the disease itself and not the cause, reintroduction of these trophic factors using viral vectors or drugs that will inhibit or activate the different molecules involved in the pro- and antiapoptotic pathways, respectively, will be an important mode of therapy for PD, Alzheimer's disease, and other neurodegenerative disorders in the future. Among these neurotrophic factors, at the present time BDNF, GDNF, and FGF2 appear to show the greatest promise.

# DYSREGULATION OF PROTEIN METABOLISM

The identification of gene mutations that are responsible for causing inherited forms of PD has expanded our focus from environmental causes of PD to the possible role of genetics in the etiology of PD. Several studies demonstrate that mutations of genes result in mutant proteins that are inefficiently catabolized by the protein removal system, the ubiquitinproteasome pathway. A defective ubiquitin-proteasome pathway can also be the result of gene defects. Failure of the ubiquitin-proteasome system to degrade a protein because it is unable to recognize the mutant protein or due to an inefficiency of the ubiquitin-proteasome system itself will result in aggregation of the mutant proteins within nigral cells and cause neurotoxicity. Evidence for both types of abnormalities has been observed in inherited forms of PD. Dysfunction of the ubiquitin-proteasome system could potentially be an important factor in the pathogenesis of PD.

#### Mutant Proteins and Inherited Forms of PD

An alanine-to-threonine substitution at codon 53 (A53T) of the gene for  $\alpha$ synuclein has been identified in several families with Italian-Greek pedigree (60). A substitution of proline for alanine at codon 30 (A30P) of the  $\alpha$ synuclein gene was also described in a family of German pedigree (61). The mRNA of  $\alpha$ -synuclein is expressed throughout the brain, but expressed at a very low intensity in the substantia nigra, and the level of expression of mRNA for  $\alpha$ -synuclein is much lower than normal in the nigral neurons of PD brains (62). The protein of  $\alpha$ -synuclein localizes to both the nucleus and the synapse, but its function is predominantly presynaptic (63).  $\alpha$ -Synuclein has structural similarities to the chaperone protein 14-3-3 (64), and together,  $\alpha$ -synuclein and 14-3-3 regulate the expression of TH (65).

Recognition that mutations of the Parkin gene are responsible for autosomal recessive-juvenile parkinsonism (AR-JP) is a major breakthrough in our understanding of the pathogenesis of PD (66). The Parkin gene maps to chromosome 6q25.2-q27 and has 12 exons coding for a 465-amino-acid protein. Mutations of the Parkin gene are the most common type noted in autosomal recessive PD (66). Parkin is an E3 ubiquitin ligase (67), an enzyme that plays an important role in the ubiquitin-proteasome protein degradation pathway. The E3 ubiquitin ligase family consists of a large number of members and among these, Parkin is the type that contains, within the same molecule, a ring finger domain that binds to ubiquitin as well as a site that recognizes and binds to the substrate (68).

 $\alpha$ -Synuclein (66), alphaSp22, which is a glycosylated isoform of  $\alpha$ -synuclein (69), CDCrel-1 (67), a synaptic vesicle–associated protein, synphilin-1 (70), Pael receptor (Parkin-associated endothelin receptor-like receptor) (71), and a G-protein–coupled transmembrane polypeptide that is expressed most intensely in the TH-positive neurons of the nigra (72) are some of the substrates that interact with Parkin.

A mutation of the gene that codes for ubiquitin carboxy-terminal hydrolase (UCH-L1) has been recognized in one family with an inherited

form of PD (73). Ubiquitin hydrolases are deubiquitinating enzymes that play a pivotal role in maintaining a steady-state level of ubiquitin by generating and recycling ubiquitin (68). A mutation of the human neurofilament M gene has also been reported in a patient with young-onset PD with a French-Canadian pedigree (74).

# Ubiquitin-Proteasome Protein Degradation Pathway and PD

Ubiquitin-proteasome-mediated protein catabolic pathway plays a major role in maintaining a viable and normal functioning cell. Dysfunction of the ubiquitin-proteasome pathway has been proposed to be involved in many neurodegenerative disorders, including Alzheimer's disease, frontotemporal dementia, Huntington's disease, and several types of malignancies (68,75). The two basic mechanisms that are involved in the catabolism of proteins by the ubiquitin-proteasome pathway are (1) the protein that needs to be degraded is tagged with ubiquitin and (2) the tagged protein is then transferred to a protease, 26S proteasome, which degrades the tagged protein into small peptides. The initial step for ubiquitination of the substrate consists of activation of the inactive form of ubiquitin by the ubiquitin-activating enzyme E1. The activated ubiquitin is then transferred to the ubiquitin carrier protein family of enzymes (ubiquitin conjugating enzyme or Ubc) E2. Among the 13 subtypes of E2, UbcH7 and UbcH8 appear to play a more prominent role in interacting with the subtype of E3 ubiquitin ligase implicated in AR-JP (66). E3 (ubiquitin ligase), consisting of a large family of ligases, is responsible for recognizing the substrate protein that is to be degraded and facilitating the transfer of activated ubiquitin from E2 so that the substrate can be tagged with ubiquitin and subsequently recognized by the proteasome. This substrate-ubiquitin complex is further polyubiquitinated by a polyubiquitinating enzyme E4 and presented to the 26S proteasome. The proteasome then degrades the tagged protein to small polypeptides, and peptidases further degrade the peptides into amino acids. Ubiquitin is released for further use by one of several deubiquitinating enzymes. The deubiquitinating enzymes play the important role of regulating the amount of ubiquitin available (Fig. 3).

Gene defects resulting in dysfunction of many of the different molecules involved in the ubiquitin-proteasome pathway have been shown to induce PD. These mutations of the genes may result in (1) mutant forms of ubiquitin-domain proteins (UDPs), which will decrease the availability of free UDPs; (2) mutant ubiquitin ligase (E3), which will result in a failure of E3 to recognize the substrate; (3) mutations of the protein substrate; or (4) a failure to deubiquitinate, which will result in a decreased supply of free ubiquitin to inadequate recycling of ubiquitin. The N-terminal region of the

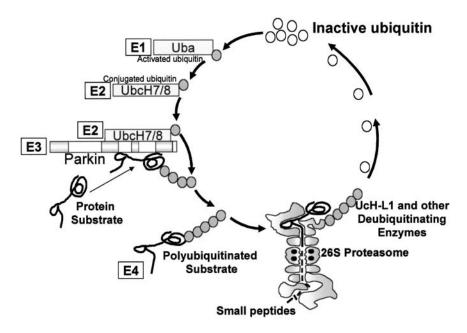


FIGURE 3 Diagrammatic representation of the Parkin-Ubiquitin proteasome pathway. See text for details. (Adapted from Refs. 66, 67, 68, 72.)

Parkin protein codes for an ubiquitin-domain protein (UDP). The UDPs have structural similarities to ubiquitin and function as proteasome adapters (76). A mutation of the ubiquitin-like domain coding regions of exons 2 and 3 of the Parkin gene alone has been observed in AR-JP (66). Deletion, duplication, and mutations of several regions of the Parkin gene have been recognized to cause AR-JP. Among these, mutations in the regions coding for ring fingers appear to be quite frequent among AR-JP patients (66). Mutation of  $\alpha$ -synuclein may result in failure of the ubiquitinating system to recognize the substrate. Even though only two members of the family were reported to have mutations of the gene of UCH-L1, resulting in only a partial suppression of the deubiquitinating enzyme UCH-L2, these two patients reinforce the concept that disturbances of the protein degradation system will lead to aggregation of protein within the neuron. Proteasomal dysfunctions have also been observed in PD (77).

# Neurochemistry of the Lewy Body

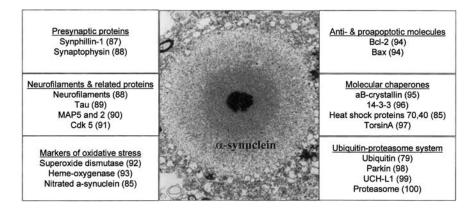
Mutated and misfolded proteins tend to aggregate. During aggregation, as commonly seen in polyglutamine repeat diseases, these fibrillary proteins may also sequester other proteins, including chaperone proteins and ubiquitin-conjugating enzymes, and further contribute to prevention of catabolism of these mutant proteins (78). The presence of the Lewy body, an example of such protein aggregation, in the substantia nigra is considered pathognomonic of PD. Electron microscopically, the Lewy body consists of a dense circular central core surrounded by neurofilaments located in the pale halo at the periphery (79). Aggregated  $\alpha$ -synuclein is a major constituent of Lewy bodies in PD. The central core as well as the peripheral halo immunostain very strongly for the full length of  $\alpha$ -synuclein (80).  $\beta$  or  $\gamma$ -Synuclein appears to aggregate in the axon terminals of hippocampal neurons of PD and diffuse Lewy body brains (81).

An overexpression of  $\alpha$ -synuclein, as demonstrated in *Drosophila* models of PD (82), is neurotoxic to nigral neurons, but this toxicity can be reversed by overexpression of two other chaperone proteins, namely HSP70 and HSP40 (83). In the presence of iron, which accumulates in the ventral mesencephalon in PD, or of aluminum, copper, or manganese ions,  $\alpha$ -synuclein appears to be insoluble (84). Increased oxidative stress can be an early step for aggregation of  $\alpha$ -synuclein (85), and in turn, aggregation of  $\alpha$ -synuclein can promote further mitochondrial dysfunction and oxidative stress (86).  $\alpha$ -Synuclein accumulation can decrease the effectiveness of proteasomal function and contribute to the accumulation of other proteins that are substrates for proteasomal degradation. Understanding the mechanisms of insolubility of  $\alpha$ -synuclein is crucial to creating new modes of therapy for PD and other synucleinopathies.

Besides  $\alpha$ -synuclein, Lewy bodies also contain several other proteins. These proteins can be broadly grouped into several types, namely (1) proteins that have presynaptic functions, (2) neurofilaments and related proteins, (3) markers of oxidative stress, (4) pro- and antiapoptotic proteins, (5) molecular chaperones, and (6) members of the ubiquitin-proteasome system (Fig. 4).

# Summary

The discovery that mutations of the  $\alpha$ -synuclein and Parkin genes cause a parkinsonian syndrome has led to a better understanding of the mechanisms with which the dopaminergic neurons of the substantia nigra handle protein degradation. The evidence indicates that in PD, as in several other synucleinopathies, the aggregation of  $\alpha$ -synuclein and ubiquitin in Lewy bodies is the result of ineffective removal of proteins by the ubiquitin-proteasomal system of the dopaminergic neurons of the substantia nigra.



**FIGURE 4** Diagrammatic representation of a Lewy body. The different proteins that are associated with Lewy bodies in PD are listed in boxes.

# MITOCHONDRIAL DAMAGE AND THE SUBSTANTIA NIGRA

The concept that mitochondrial dysfunction can cause a parkinsonian syndrome came into focus with the observation that MPTP induced PD in "frozen" addicts (101–103). Mitochondrion is the major source of cellular energy. Each cell has thousands of mitochondria throughout the cytoplasm. In addition to generating most of the energy in the form of adenosine triphosphate (ATP) required by the cell through the oxidative phosphorylation system (OXPHOS), mitochondria also generate and remove free radicals and play the central role in initiating many of the key steps for apoptosis (104). Mitochondrial dysfunctions are now recognized to be the major cause of nigral degeneration in experimental models of PD (105,106) and possibly even in idiopathic PD.

#### **Electron Transfer Chain Dysfunction**

The electron transfer chain (ETC) is an important component of the OXPHOS system. The respiratory complexes I, II, III, IV, and V are located within the inner membrane of the mitochondria and play a critical role in transferring electrons from different sources within the mitochondria. Dysfunction of these respiratory complexes will lead to significant loss in the generation of stored energy in the form of ATP as well as increased oxidative damage due to accumulation of reactive oxygen species.

The MPTP model of PD clearly suggests that inhibition of the ETC, especially at the Complex I level, is toxic to nigral neurons (107). There is a

significant decrease in the levels of complex I in the nigral neurons of PD (108,109). Among the different diseases of the basal ganglia, the deficiency of Complex I in the nigra may be specific to PD (110). Complex I deficiency is not restricted to the brain, but is also found in the skeletal muscle, platelets, fibroblasts, and lymphocytes (111) in PD. It is important to recognize that in the two most commonly used animal models of PD, 6-OHDA– and MPTP-induced models, the toxins decrease the efficiency of Complex I (105,106).

Chronic injections of a commonly used pesticide, rotenone, cause selective degeneration of the nigrostriatal system and result in aggregation of ubiquitin and  $\alpha$ -synuclein (112). This is important in understanding the etiology of PD. Rotenone, a toxin that is used to kill fish in ponds, is a mitochondrial toxin, which easily crosses the blood-brain barrier and inhibits Complex I. The clinical syndrome that results with chronic rotenone administration has significant similarities to human PD, including hypokinesia, stooped posture, and tremor. A dysfunction of Complex III has also been suggested to occur in PD (113). Recent studies suggest that nitric oxide (NO) can inhibit Complex IV, and this inhibition may accentuate the toxic effects of methyl-4-phenylpyridium(MPP+) on Complex I (114).

# Free Radicals and Mitochondrial Toxicity

Mitochondria are the major source of energy production for the cell and the major site of utilization of cellular oxygen. During the synthesis of stored energy in the form of ATP, mitochondria produce several reactive oxygen species (ROS). It is estimated that 2–4% of the utilized oxygen is converted into ROS. ROS consists of superoxide anions, hydroxyl radicals, and hydrogen peroxide. The majority of the ROS produced in a cell is derived from the mitochondria. ROS are produced at Complex I and Complex III of the ETC system (115).

ROS are toxic to neurons. An increased level of ROS within the mitochondria can decrease the efficiency of Complex I and the ETC system. This will decrease energy production and cause further accumulation of free radicals, mtDNA damage, and production of additional free radicals through Fenton reaction. All of these events may induce aging of the cell and mitochondria-mediated apoptotic mechanisms of cell death (115). An increased accumulation of ROS can occur either due to increased synthesis of ROS or because of failure or a decreased level of removal by the glutathione system. Evidence for both increased production as well as decreased clearance of these free radicals has been observed in PD.

The different metabolic breakdown pathways of levodopa may be an important source of an increased production of free radicals. There is controversy regarding the neurotoxic vs. neuroprotective role of levodopa in PD (116–118). There is extensive literature to suggest that, at least in experimental conditions, especially in conditions using cell culture techniques, levodopa can produce free radicals and can destroy the cells in culture. In the presence of iron, there is an even greater increase in the levels of free radicals synthesized (119,120). Extensive review of this issue suggests that the evidence supporting the neurotoxic role of levodopa is insufficient in patients with PD (118). In fact, the cytotoxic effects of dopamine may be an artifact of cell culture technique (121), and the neurotoxic effects of accumulated iron may be due to its ability to promote aggregation of  $\alpha$ -synuclein (84) and may contribute to Lewy body pathology.

An increased level of free radicals may be from sources other than the mitochondria. NO may be one such source of free radicals (122). An increased production of nitric oxide synthase, as noted in MPTP models of PD (123), can lead to increased levels of NO, which in turn results in an increased synthesis and accumulation of free radicals within the nigral cells. NO may play an important role in inducing nigral degeneration in amphetamine and MPTP-induced PD. Increased NO levels can lead to increased formation of peroxynitrite, a potent oxidant, and peroxynitrite may interact with dopamine autooxidant products and neuromelanin (124,125) and can cause free radical mediated nigral neurotoxicity.

The presence of a defective free radical removal system has been established in PD. ROS is removed by superoxide dismutase (SOD), catalase, and glutathione peroxidase. SOD facilitates the conversion of superoxide to hydrogen peroxide, and catalase and glutathione peroxidase convert hydrogen peroxide to water (115). Decreased levels of glutathione, glutathione peroxidase, and catalase have been observed in PD (126,127). In fact, glutathione depletion may be one of the earliest events in the evolution of mitochondrial dysfunctions in PD (128). Decreased glutathione may actually cause a selective inhibition of Complex I and result in an inefficient ETC system (129,130).

# mtDNA Defects

Mitochondria's own genome (mtDNA) is localized in the matrix compartment. mtDNA consists of a 16.5 kb molecule coding for a total of 37 genes, 13 of which code for the oxidative phosphorylation system (OXPHOS), 2 for ribosomal RNAs (12S and 16S rRNA), and 22 for transfer RNAs (tRNA), molecules that are necessary for the translation of mtDNA structural genes (104,131). Mutations of mtDNA increase with age. Cybrid studies have suggested that the Complex I deficiency noted in idiopathic PD may be due to mtDNA aberrations. Transmitochondridal cybrid lines, using mitochondria of platelets from PD patients, show several features of oxidative stress, increased vulnerability to MPP+, and an increased expression of apoptotic molecules (132,133). More recently, the presence of tRNA mutations have been observed in histologically confirmed PD patients (134).

# Summary

This evidence suggests that toxins in the environment and toxins generated intrinsically can result in dysfunction of nigral mitochondria and lead to nigral degeneration and the induction of a slowly progressive syndrome similar to PD.

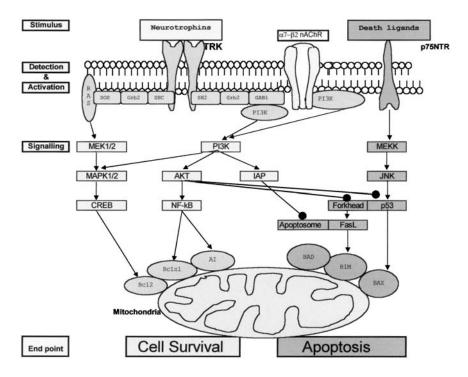
# **NIGRA AND APOPTOSIS**

There is a significant controversy about whether apoptotic mechanisms play a role in the death of substantia nigra neurons in PD (135,136). However, experimental evidence of several well-recognized markers of apoptosis has been demonstrated in the nigral cells of both experimental models of PD and human PD (137).

Apoptosis, an established mechanism of cell death during embryogenesis and brain maturation, is characterized by a well-defined stereotypical pattern of neurochemical and morphological changes (138,139). The process involves caspase-induced cleaving of DNA and numerous other intracellular polypeptides, which ultimately leads to cellular death. Activation of caspases is a key triggering event for apoptosis. Caspases may be activated by at least by two different mechanisms: an extrinsic system which involves stimulation of death receptors by extracellular "death ligands" (139) or an intrinsic pathway, which requires the activation of the mitochondrial pathway. Death ligands bind to members of TNF receptor gene superfamily of death receptors, namely Fas, TNFR1, DR3, DR4, and DR5 (38), and cause apoptosis. Once stimulated, the death receptors activate the initiator caspase 9, and this in turn activates effector caspases 3 and 6 and executes apoptosis.

# Mitochondrial Apoptotic Pathways and the Substantia Nigra

Many apoptotic and antiapoptotic factors converge upon the mitochondria. The expression of many of these factors is dependent on the activation of neurotrophin and *trk*-mediated antiapoptotic or the p75<sup>NTR</sup>-mediated apoptotic molecular cascades (Fig. 5). The members of the Bcl-2 family, namely Bcl-2, Bcl-xl, and Bcl-w, are antiapoptotic. Bax, Bak, and another group of polypeptides that includes Bid, Bad, Bim, and Bik are proapoptotic (139) (Figs. 4 and 5). Activation of caspases through the mitochondrial pathway is mediated by proapoptotic molecules facilitating the release of cytochrome *c* from the mitochondria as well as apoptotic-inducing factor (AIF) and endonuclease G, two other molecules released from the mitochondria that participate in apoptosis. Release of cytochrome *c* into the cytoplasm triggers activation of the effector caspases 3 and 6 and results in the destruction of the cytoplasm. The AIF translocates to the nucleus and contributes to DNA fragmentation and chromatin destruction and ultimately to cell death (139).



**FIGURE 5** The neurotrophin-mediated pathways influencing the expression of different proapoptotic and antiapoptotic molecules on the mitochondria. (Adapted from Refs. 35,139,158.)

Bax, the proapoptotic protein, is expressed ubiquitously in normal and degenerating neurons of human nigra, but the numbers of Bax-positive neurons are significantly higher among the melanized degenerating nigral neurons in PD (140,141). p53-deficient mice were resistant to MPP+-induced neurotoxicity of dopaminergic neurons of nigra (142). Bax ablation prevents the MPTP-induced mouse model of PD (143). Overexpression of Bcl-2 protects catecholaminergic neurons from MPP+ and 6-OHDA toxicity of PC12 cells and neurons (144,145).

The effector caspase, caspase 3, is activated in 6-OHDA models of PD (146–149). Similarly, MPTP-induced nigral neurotoxicity has also been shown to be apoptotic. MPP+ also induces significant elevations of levels of caspase 3, caspase 8, and caspase 1 in the nigral neurons of mice, and inhibition of caspase activity prevents MPP+ neurotoxicity (150). Caspase 1 and 3 activities are increased in human nigral dopaminergic neurons of PD (151–153).

TUNEL-positive cells, an indicator of DNA fragmentation from apoptosis, have been noted in the MPTP-induced degeneration of nigral neurons and in melanized dopaminergic neurons of human nigra in PD (154–156). These and other studies strongly suggest that activation of the molecular cascades of proapoptotic pathways plays an important role in the death of substantia nigra neurons in PD.

# Death Receptors and the Substantia Nigra

Apoptosis can also be induced by activation of several receptors of the TNF receptor gene superfamily. When "death ligands" bind to the receptors, the cells trigger several molecules that instruct the cells to self-destruct. So far, the ligands that directly activate the death receptors in PD have not been identified. TNF-receptor1, a receptor that contains the "death domain," is expressed in the nigral dopaminergic neurons and the glial cells that express this receptor. These are higher in the substantia nigra of PD than in controls (157).

# Summary

The evidence for mitochondrial apoptotic pathways playing an important role in the death of dopaminergic neurons in PD is being established. The possible role of the receptors with death domain in the pathogenesis of PD and the role of increased levels of cytokines that have the potential to stimulate these death receptors is just beginning to be explored.

# CONCLUSION

From the evidence discussed above, it is clear that both environmental and genetic factors can cause a parkinsonian syndrome that is similar to idiopathic PD. Environmental toxins induced mitochondrial dysfunction might be the most common cause of PD. In this regard, the observation that chronic administration of rotenone, a common toxin, can lead to a neurochemical and clinical syndrome that has significant similarities to that of idiopathic PD is a seminal one. This study certainly reinforces the prevailing epidemiological evidence that environmental toxins may cause PD.

However, based on the pathogenesis of familial PD, it is also recognized that dysfunction of the ubiquitin-proteasome system can lead to protein aggregation and resultant toxicity to nigral neurons. Accumulation and aggregation of proteins because of abnormal folding of proteins or inefficiency of molecular chaperones or proteasomal functions may occur even in the absence of any gene defects or toxins. Such mechanisms may play a role in aggregation of proteins in non-familial forms of Alzheimer's, prion and motor neuron diseases (68,75,78).

The survival of a neuron may also be dependant on a delicate balance between the neurotrophin and *trk* receptor–mediated antiapoptotic pathway and the proapoptotic pathway mediated by  $p75^{NTR}$  and other death receptors (139,158). As of yet, there is no direct evidence to support the hypothesis that withdrawal of growth factors or stimulation of the death receptors and accessing the direct pathway to apoptosis exists in PD. However, the role of a decreased level of growth factors as well as increased levels of cytokines observed in the dopaminergic neurons of the substantia nigra in PD remains to be explored.

As the understanding of the trophic factors that influence normal differentiation and maturation of nigral dopaminergic neurons is expanding, the possibility that lack of or reduced influence of these ontogenic trophic factors may somehow result in either a decreased number or defective dopamine neurons in adulthood exists. Such a decrease in the number or efficiency of dopamine neurons may be a risk factor for developing PD later in life.

While many of these concepts about multiple etiologies for PD are nothing more than hypothetical at present, the knowledge derived from modern experimental approaches will certainly allow us to enter into a new and exciting phase of diagnosing these patients early in the course of the disease and treat them with molecules that will either slow the progression of PD or, hopefully, even stop the progression of the disease.

# ACKNOWLEDGMENTS

Supported by the Carl Baldridge Parkinson's Disease Research Fund. Dedicated to my mentors Drs. B. Ramamurthi, T. Umeshraya Pai, C. H. Narayanan, Malcolm Carpenter, Stanley Fahn, Joseph Chusid, Hyman Donnenfeld, and George Uhl.

# REFERENCES

- R Hassler. Zur Normalantomie de Substantia nigra. J Psychol Neurol 48:1–55, 1937.
- R. Hassler. Zur Pathologie der Paralysis agitans und des postencephalitischen Parkinsonismus. J. psychol. Neurology 48:387–455, 1938.
- 3. JM Fearnley, AJ Lees. Ageing and Parkinson's disease: substantia nigra regional selectivity. Brain 114:2283–2301, 1991.
- WR Gibb, AJ Lees. Anatomy, pigmentation, ventral and dorsal subpopulations of the substantia nigra, and differential cell death in Parkinson's disease. J Neurol Neurosurg Psychiatry 54:388–396, 1991.
- 5. Parkinson Study Group. Dopamine transporter brain imaging to assess the effects of pramipexole vs levodopa on Parkinson disease progression. JAMA 287:1653–1661,2002.
- 6. YA Barde. Trophic factors and neuronal survival. Neuron 2:1525–1534,1989.
- 7. MV Sofroniew, CL Howe, WC Mobley. Nerve growth factor signaling, neuroprotection, and neural repair. Annu Rev Neurosci 24:1217–1281, 2001.
- 8. B Connor, M Dragunow. The role of neuronal growth factors in neurodegenerative disorders of the human brain. Brain Res Brain Res Rev 27:1–39, 1998.
- M Hynes, JA Porter, C Chiang, D Chang, M Tessier-Lavigne, PA Beachy, A Rosenthal. Induction of midbrain dopaminergic neurons by sonic hedgehog. Neuron 15:35–44, 1995.
- 10. J Briscoe, J Erickson. Specification of neuronal fates in the ventral neural tube. Curr Opin Neurobiol 11:43–49, 2001.
- 11. K Tsuboi, CW Shults. Intrastriatal injection of sonic hedgehog reduces behavioral impairment in a rat model of Parkinson's disease. Exp Neurol 173:95–104, 2002.
- O Saucedo-Cardenas, JD Quintana-Hau, WD Le, MP Smidt, JJ Cox, F De Mayo, JP Burbach, OM Conneely. Nurrl is essential for the induction of the dopaminergic phenotype and the survival of ventral mesencephalic late dopaminergic precursor neurons. Proc Natl Acad Sci USA 95:4013–4018, 1998.
- C Backman, T Perlmann, A Wallen, BJ Hoffer, M Morales. A selective group of dopaminergic neurons express Nurrl in the adult mouse brain. Brain Res 851:125–132, 1999.
- RH Zetterstrom, L Solomin, L Jansson, BJ Hoffer, L Olson, T Perlmann. Dopamine neuron agenesis in Nurrl-deficient mice. Science 276:248–250, 1997.

- SO Castillo, JS Baffi, M Palkovits, DS Goldstein, IJ Kopin, J Witta, MA Magnuson, VM Nikodem. Dopamine biosynthesis is selectively abolished in substantia nigra/ventral tegmental area but not in hypothalamic neurons in mice with targeted disruption of the Nurrl gene. Mol Cell Neurosci 11:36–46, 1998.
- W Le, OM Conneely, L Zou, Y He, O Saucedo-Cardenas, J Jankovic, DR Mosier, SH Appel. Selective agenesis of mesencephalic dopaminergic neurons in Nurrl-deficient mice. Exp Neurol 159:451–458, 1999.
- K Sakurada, M Ohshima-Sakurada, TD Palmer, FH Gage. Nurrl, an orphan nuclear receptor, is a transcriptional activator of endogenous tyrosine hydroxylase in neural progenitor cells derived from the adult brain. Development 126:4017–4026, 1999.
- JS Baffi, M Palkovits, SO Castillo, E Mezey, VM Nikodem. Differential expression of tyrosine hydroxylase in catecholaminergic neurons of neonatal wild-type and Nurrl-deficient mice. Neuroscience 93:631–642, 1999.
- 19. T Iwawaki, K Kohno, K Kobayashi. Identification of a potential nurrl response element that activates the tyrosine hydroxylase gene promoter in cultured cells. Biochem Biophys Res Commun 274:590–595, 2000.
- 20. P Sacchetti, TR Mitchell, JG Granneman, MJ Bannon. Nurrl enhances transcription of the human dopamine transporter gene through a novel mechanism. J Neurochem 76:1565–1572, 2001.
- RH Zetterstrom, R Williams, T Perlmann, L Olson. Cellular expression of the immediate early transcription factors Nurrl and NGFI-B suggests a gene regulatory role in several brain regions including the nigrostriatal dopamine system. Brain Res Mol Brain Res 41:111–120, 1996.
- 22. W Le, OM Conneely, Y He, J Jankovic, SH Appel. Reduced Nurrl expression increases the vulnerability of mesencephalic dopamine neurons to MPTP-induced injury. J Neurochem 73:2218–2221, 1999.
- 23. PY Xu, R Liang, J Jankovic, C Hunter, YX Zeng, T Ashizawa, D Lai, WD Le. Association of homozygous 7048G7049 variant in the intron six of Nurrl gene with Parkinson's disease. Neurology 58:881–884, 2002.
- MP Smidt, HS van Schaick, C Lanctot, JJ Tremblay, JJ Cox, AA van der Kleij, G Wolterink, J Drouin, JP Burbach. A homeodomain gene Ptx3 has highly restricted brain expression in mesencephalic dopaminergic neurons. Proc Natl Acad Sci USA 94:13305–13310, 1997.
- M Lebel, Y Gauthier, A Moreau, J Drouin. Pitx3 activates mouse tyrosine hydroxylase promoter via a high-affinity binding site. J Neurochem 77:558– 567, 2001.
- M Fujita, S Shimada, T Nishimura, GR Uhl, M Tohyama. Ontogeny of dopamine transporter mRNA expression in the rat brain. Brain Res Mol Brain Res 19:222–226, 1993.
- 27. SR Hansson, BJ Hoffman, E Mezey. Ontogeny of vesicular monoamine transporter mRNAs VMAT1 and VMAT2. I. The developing rat central nervous system. Brain Res Dev Brain Res 110:135–158, 1998.

- C Perrone-Capano, P Da Pozzo, U di Porzio. Epigenetic cues in midbrain dopaminergic neuron development. Neurosci Biobehav Rev 24:119–124, 2000.
- 29. DB Ramsden, RB Parsons, SL Ho, RH Waring. The aetiology of idiopathic Parkinson's disease. Mol Pathol 54:369–380, 2001.
- MP Smidt, CH Asbreuk, JJ Cox, H Chen, RL Johnson, JP Burbach. A second independent pathway for development of mesencephalic dopaminergic neurons requires Lmxlb. Nat Neurosci 3:337–341, 2000.
- LM Farkas, K Krieglstein. Heparin-binding epidermal growth factor-like growth factor (HB-EGF) regulates survival of midbrain dopaminergic neurons. J Neural Transm 109:267–277, 2002.
- HH Simon, H Saueressig, W Wurst, MD Goulding, DD O'Leary. Fate of midbrain dopaminergic neurons controlled by the engrailed genes. J Neurosci 21:3126–3134, 2001.
- GJ Siegel, NB Chauhan. Neurotrophic factors in Alzheimer's and Parkinson's disease brain. Brain Res Brain Res Rev 33:199–227, 2000.
- 34. DR Kaplan, FD Miller. Signal transduction by the neurotrophin receptors. Curr Opin Cell Biol 9:213–221, 1997.
- DR Kaplan, FD Miller. Neurotrophin signal transduction in the nervous system. Curr Opin Neurobiol 10:381–391, 2000.
- 36. Bibel, E Hoppe, YA Barde. Biochemical and functional interactions between the neurotrophin receptors trk and p75NTR. Embo J 18:616–622, 1999.
- G Dechant. Molecular interactions between neurotrophin receptors. Cell Tissue Res 305:229–238, 2001.
- A Ashkenazi, VM Dixit. Death receptors: signaling and modulation. Science 281:1305–1308, 1998.
- T Nishio, S Furukawa, I Akiguchi, N Sunohara. Medial nigral dopamine neurons have rich neurotrophin support in humans. Neuroreport 9:2847– 2851, 1998.
- 40. EJ Mufson, JS Kroin, TJ Sendera, T Sobreviela. Distribution and retrograde transport of trophic factors in the central nervous system: functional implications for the treatment of neurodegenerative diseases. Prog Neurobiol 57:451–484, 1999.
- M Mogi, A Togari, T Kondo, Y Mizuno, O Komure, S Kuno, H Ichinose, T Nagatsu. Brain-derived growth factor and nerve growth factor concentrations are decreased in the substantia nigra in Parkinson's disease. Neurosci Lett 270:45–48, 1999.
- 42. K Parain, MG Murer, Q Yan, B Faucheux, Y Agid, E Hirsch, R Raisman-Vozari. Reduced expression of brain-derived neurotrophic factor protein in Parkinson's disease substantia nigra. Neuroreport 10:557–561, 1999.
- 43. DM Frim, TA Uhler, WR Galpern, MF Beal, XO Breakefield, O Isacson. Implanted fibroblasts genetically engineered to produce brain-derived neurotrophic factor prevent 1-methyl-4-phenylpyridinium toxicity to dopaminergic neurons in the rat. Proc Natl Acad Sci USA 91:5104–5108, 1994.

- 44. NB Chauhan, GJ Siegel, JM Lee. Depletion of glial cell line-derived neurotrophic factor in substantia nigra neurons of Parkinson's disease brain. J Chem Neuroanat 21:277–288, 2001.
- 45. CW Shults, T Kimber, D Martin. Intrastriatal injection of GDNF attenuates the effects of 6-hydroxydopamine. Neuroreport 7:627–631, 1996.
- 46. JH Kordower, ME Emborg, J Bloch, SY Ma, Y Chu, L Leventhal, J McBride, EY Chen, S Palfi, BZ Roitberg, WD Brown, JE Holden, R Pyzalski, MD Taylor, P Carvey, Z Ling, D Trono, P Hantraye, N Deglon, P Aebischer. Neurodegeneration prevented by lentiviral vector delivery of GDNF in primate models of Parkinson's disease. Science 290:767–773, 2000.
- L Feng, CY Wang, H Jiang, C Oho, M Dugich-Djordjevic, L Mei, B Lu. Differential signaling of glial cell line-derived neurothrophic factor and brainderived neurotrophic factor in cultured ventral mesencephalic neurons. Neuroscience 93:265–273, 1999.
- 48. NJ Rothwell. Annual review prize lecture cytokines—killers in the brain? J Physiol 514:3–17, 1999.
- 49. R Martin, CS Sturzebecher, HF McFarland. Immunotherapy of multiple sclerosis: Where are we? Where should we go? Nat Immunol 2:785–788, 2001.
- M Mogi, M Harada, T Kondo, P Riederer, H Inagaki, M Minami, T Nagatsu. Interleukin-1 beta, interleukin-6, epidermal growth factor and transforming growth factor-alpha are elevated in the brain from parkinsonian patients. Neurosci Lett 180:147–150, 1994.
- T Nagatsu, M Mogi, H Ichinose, A Togari. Cytokines in Parkinson's disease. J Neural Transm (suppl)58:143–151, 2000.
- 52. A Cintra, YH Cao, C Oellig, B Tinner, F Bortolotti, M Goldstein, RF Pettersson, K Fuxe. Basic FGF is present in dopaminergic neurons of the ventral midbrain of the rat. Neuroreport 2:597–600, 1991.
- I Tooyama, T Kawamata, D Walker, T Yamada, K Hanai, H Kimura, M Iwane, K Igarashi, EG McGeer, PL McGeer. Loss of basic fibroblast growth factor in substantia nigra neurons in Parkinson's disease. Neurology 43:372– 376, 1993.
- 54. I Tooyama, EG McGeer, T Kawamata, H Kimura, PL McGeer. Retention of basic fibroblast growth factor immunoreactivity in dopaminergic neurons of the substantia nigra during normal aging in humans contrasts with loss in Parkinson's disease. Brain Res 656:165–168, 1994.
- 55. T Kihara, S Shimohama, H Sawada, K Honda, T Nakamizo, H Shibasaki, T Kume, A Akaike. alpha 7 nicotinic receptor transduces signals to phosphatidylinositol 3-kinase to block A beta-amyloid-induced neurotoxicity. J Biol Chem 276:13541–13546, 2001.
- P Marin, M Maus, S Desagher, J Glowinski, J Premont. Nicotine protects cultured striatal neurones against N-methyl-D-aspartate receptor-mediated neurotoxicity. Neuroreport 5:1977–1980, 1994.
- 57. R Maggio, M Riva, F Vaglini, F Fornai, R Molteni, M Armogida, G Racagni, GU Corsini. Nicotine prevents experimental parkinsonism in rodents and

induces striatal increase of neurotrophic factors. J Neurochem 71:2439–2446, 1998.

- RE Ryan, SA Ross, J Drago, RE Loiacono. Dose-related neuroprotective effects of chronic nicotine in 6-hydroxydopamine treated rats, and loss of neuroprotection in alpha4 nicotinic receptor subunit knockout mice. Br J Pharmacol 132:1650–1656, 2001.
- 59. M Quik, DA Di Monte. Nicotine administration reduces striatal MPP+levels in mice. Brain Res 917:219–224, 2001.
- 60. MH Polymeropoulos, C Lavedan, E Leroy, SE Ide, A Dehejia, A Dutra, B Pike, H Root, J Rubenstein, R Boyer, ES Stenroos, S Chandrasekharappa, A Athanassiadou, T Papapetropoulos, WG Johnson, AM Lazzarini, RC Duvoisin, G Di Iorio, LI Golbe, RL Nussbaum. Mutation in the alphasynuclein gene identified in families with Parkinson's disease. Science 276:2045–2047, 1997
- R Kruger, W Kuhn, T Muller, D Woitalla, M Graeber, S Kosel, H Przuntek, JT Epplen, L Schols, O Riess. Ala30Pro mutation in the gene encoding alphasynuclein in Parkinson's disease. Nat Genet 18:106–108, 1998.
- 62. M Neystat, T Lynch, S Przedborski, N Kholodilov, M Rzhetskaya, RE Burke. Alpha-synuclein expression in substantia nigra and cortex in Parkinson's disease. Mov Disord 14:417–422, 1999.
- NB Cole, DD Murphy. The cell biology of alpha-synuclein: a sticky problem? Neuromolecular Med 1:95–109, 2002. LA Hansen, M Mallory, JQ Trojanowski, D Galasko, E Masliah. Altered expression of the synuclein family mRNA in Lewy body and Alzheimer's disease. Brain Res 914:48–56, 2001.
- 64. N Ostrerova, L Petrucelli, M Farrer, N Mehta, P Choi, J Hardy, B Wolozin. Alpha-synuclein shares physical and functional homology with 14-3-3 proteins. J Neurosci 19:5782–5791, 1999.
- RG Perez, JC Waymire, E Lin, JJ Liu, F Guo, MJ Zigmond. A role for alphasynuclein in the regulation of dopamine biosynthesis. J Neurosci 22:3090– 3099, 2002.
- 66. K Tanaka, T Suzuki, T Chiba, H Shimura, N Hattori, Y Mizuno. Parkin is linked to the ubiquitin pathway. J Mol Med 79:482–494, 2001.
- Y Zhang, J Gao, KK Chung, H Huang, VL Dawson, TM Dawson. Parkin functions as an E2-dependent ubiquitin- protein ligase and promotes the degradation of the synaptic vesicle-associated protein, CDCrel-1. Proc Natl Acad Sci USA 97:13354–13359, 2000.
- 68. MH Glickman, A Ciechanover. The ubiquitin-proteasome proteolytic pathway: destruction for the sake of construction. Physiol Rev 82:373–428, 2002.
- H Shimura, MG Schlossmacher, N Hattori, MP Frosch, A Trockenbacher, R Schneider, Y Mizuno, KS Kosik, DJ Selkoe. Ubiquitination of a new form of alpha-synuclein by parkin from human brain: implications for Parkinson's disease. Science 293:263–269, 2001.
- 70. KK Chung, Y Zhang, KL Lim, Y Tanaka, H Huang, J Gao, CA Ross, VL Dawson, TM Dawson. Parkin ubiquitinates the alpha-synuclein-interacting

protein, synphilin-1: implications for Lewy-body formation in Parkinson disease. Nat Med 7:1144-1150, 2001.

- 71. Y Imai, M Soda, H Inoue, N Hattori, Y Mizuno, R Takahashi. An unfolded putative transmembrane polypeptide, which can lead to endoplasmic reticulum stress, is a substrate of Parkin. Cell 105:891–902, 2001.
- 72. BI Giasson, VM Lee. Parkin and the molecular pathways of Parkinson's disease. Neuron 31:885–888, 2001.
- 73. E Leroy, R Boyer, G Auburger, B Leube, G Ulm, E Mezey, G Harta, MJ Brownstein, S Jonnalagada, T Chernova, A Dehejia, C Lavedan, T Gasser, PJ Steinbach, KD Wilkinson, MH Polymeropoulos. The ubiquitin pathway in Parkinson's disease. Nature 395:451–452, 1998.
- 74. C Lavedan, S Buchholtz, RL Nussbaum, RL Albin, MH Polymeropoulos. A mutation in the human neurofilament M gene in Parkinson's disease that suggests a role for the cytoskeleton in neuronal degeneration. Neurosci Lett 322:57–61, 2002.
- 75. R Layfield, A Alban, RJ Mayer, J Lowe. The ubiquitin protein catabolic disorders. Neuropathol Appl Neurobiol 27:171–179, 2001.
- S Jentsch, G Pyrowolakis. Ubiquitin and its kin: How close are the family ties? Trends Cell Biol 10:335–342, 2000.
- 77. KS McNaught, P Jenner. Proteasomal function is impaired in substantia nigra in Parkinson's disease. Neurosci Lett 297:191–194, 2001.
- MD Kaytor, ST Warren. Aberrant protein deposition and neurological disease. J Biol Chem 274:37507–37510, 1999.
- 79. H Takahashi, K Wakabayashi. The cellular pathology of Parkinson's disease. Neuropathology 21:315–322, 2001.
- MG Spillantini, RA Crowther, R Jakes, M Hasegawa, M Goedert. alphasynuclein in filamentous inclusions of Lewy bodies from Parkinson's disease and dementia with lewy bodies. Proc Natl Acad Sci USA 95:6469–6473, 1998.
- JE Galvin, K Uryu, VM Lee, JQ Trojanowski. Axon pathology in Parkinson's disease and Lewy body dementia hippocampus contains alpha-, beta-, and gamma-synuclein. Proc Natl Acad Sci USA 96:13450–13455, 1999.
- 82. MB Feany and WW Bender. A Drosophila model of Parkinson's disease. Nature 404:394–398, 2000.
- PK Auluck, HY Chan, JQ Trojanowski, VM Lee, NM Bonini. Chaperone suppression of alpha-synuclein toxicity in a Drosophila model for Parkinson's disease. Science 295:865–868, 2002.
- VN Uversky, J Li, AL Fink. Metal-triggered structural transformations, aggregation, and fibrillation of human alpha-synuclein. A possible molecular NK between Parkinson's disease and heavy metal exposure. J Biol Chem 276:44284–44296, 2001.
- 85. BI Giasson, JE Duda, IV Murray, Q Chen, JM Souza, HI Hurtig, H Ischiropoulos, JQ Trojanowski, VM Lee. Oxidative damage linked to neurodegeneration by selective alpha-synuclein nitration in synucleinopathy lesions. Science 290:985–989, 2000.

- LJ Hsu, Y Sagara, A Arroyo, E Rockenstein, A Sisk, M Mallory, J Wong, T Takenouchi, M Hashimoto, E Masliah. alpha-synuclein promotes mitochondrial deficit and oxidative stress. Am J Pathol 157:401–410, 2000.
- S Engelender, Z Kaminsky, X Guo, AH Sharp, RK Amaravi, JJ Kleiderlein, RL Margolis, JC Troncoso, AA Lanahan, PF Worley, VL Dawson, TM Dawson, CA Ross. Synphilin-1 associates with alpha-synuclein and promotes the formation of cytosolic inclusions. Nat Genet 22:110–114, 1999.
- A Takeda, M Mallory, M Sundsmo, W Honer, L Hansen, E Masliah. Abnormal accumulation of NACP/alpha-synuclein in neurodegenerative disorders. Am J Pathol 152:367–372, 1998.
- K Arima, S Hirai, N Sunohara, K Aoto, Y Izumiyama, K Ueda, K Ikeda, M Kawai. Cellular co-localization of phosphorylated tau- and NACP/alphasynuclein-epitopes in lewy bodies in sporadic Parkinson's disease and in dementia with Lewy bodies. Brain Res 843:53–61, 1999.
- 90. WP Gai, PC Blumbergs and WW Blessing. Microtubule-associated protein 5 is a component of Lewy bodies and Lewy neurites in the brainstem and forebrain regions affected in Parkinson's disease. Acta Neuropathol (Berl) 91:78–81, 1996.
- 91. S Nakamura, Y Kawamoto, S Nakano, I Akiguchi and J Kimura. p35nck5a and cyclin-dependent kinase 5 colocalize in Lewy bodies of brains with Parkinson's disease. Acta Neuropathol (Berl) 94:153–157, 1997.
- 92. K Nishiyama, S Murayama, J Shimizu, Y Ohya, S Kwak, K Asayama, I Kanazawa. Cu/Zn superoxide dismutase-like immunoreactivity is present in Lewy bodies from Parkinson disease: a light and electron microscopic immunocytochemical study. Acta Neuropathol (Berl) 89:471–474, 1995.
- 93. HM Schipper, A Liberman, EG Stopa. Neural heme oxygenase-1 expression in idiopathic Parkinson's disease. Exp Neurol 150:60–68, 1998.
- A Tortosa, E Lopez, I Ferrer. Bcl-2 and Bax proteins in Lewy bodies from patients with Parkinson's disease and diffuse Lewy body disease. Neurosci Lett 238:78–80, 1997.
- 95. J Lowe, H McDermott, I Pike, I Spendlove, M Landon, RJ Mayer. alpha B crystallin expression in non-lenticular tissues and selective presence in ubiquitinated inclusion bodies in human disease. J Pathol 166:61–68, 1992.
- 96. Y Kawamoto, I Akiguchi, S Nakamura, Y Honjyo, H Shibasaki, H Budka. 14-3-3 proteins in Lewy bodies in Parkinson disease and diffuse Lewy body disease brains. J Neuropathol Exp Neurol 61:245–253, 2002.
- P Shashidharan, PF Good, A Hsu, DP Perl, MF Brin, CW Olanow. TorsinA accumulation in Lewy bodies in sporadic Parkinson's disease. Brain Res 877:379–381, 2000.
- MG Schlossmacher, MP Frosch, WP Gai, M Medina, N Sharma, L Forno, T Ochiishi, H Shimura, R Sharon, N Hattori, JW Langston, Y Mizuno, BT Hyman, DJ Selkoe, KS Kosik. Parkin localizes to the Lewy bodies of Parkinson disease and dementia with Lewy bodies. Am J Pathol 160:1655– 1667, 2002.

- J Lowe, H McDermott, M Landon, RJ Mayer, KD Wilkinson. Ubiquitin carboxyl-terminal hydrolase (PGP 9.5) is selectively present in ubiquitinated inclusion bodies characteristic of human neurodegenerative diseases. J Pathol 161:153–160, 1990.
- 100. K Ii, H Ito, K Tanaka and A Hirano. Immunocytochemical co-localization of the proteasome in ubiquitinated structures in neurodegenerative diseases and the elderly. J Neuropathol Exp Neurol 56:125–131, 1997.
- GC Davis, AC Williams, SP Markey, MH Ebert, ED Caine, CM Reichert, IJ Kopin. Chronic Parkinsonism secondary to intravenous injection of meperidine analogues. Psychiatry Res 1:249–254, 1979.
- JW Langston, P Ballard, JW Tetrud, I Irwin. Chronic Parkinsonism in humans due to a product of meperidine-analog synthesis. Science 219:979– 980, 1983.
- 103. JW Langston. The Case of the Frozen Addicts. Santa Rosa, CA: Vintage Publications, 1996.
- DC Wallace. Mitochondrial diseases in man and mouse. Science 283:1482– 1488, 1999.
- 105. HS Chun, GE Gibson, LA DeGiorgio, H Zhang, VJ Kidd, JH Son. Dopaminergic cell death induced by MPP(+), oxidant and specific neurotoxicants shares the common molecular mechanism. J Neurochem 76:1010–1021, 2001.
- R Betarbet, TB Sherer, JT Greenamyre. Animal models of Parkinson's disease. Bioessays 24:308–318, 2002.
- I Vyas, RE Heikkila, WJ Nicklas. Studies on the neurotoxicity of 1-methyl-4phenyl-1,2,3,6-tetrahydropyridine: inhibition of NAD-linked substrate oxidation by its metabolite, 1-methyl-4-phenylpyridinium. J Neurochem 46:1501– 1507, 1986.
- 108. Y Mizuno, S Ohta, M Tanaka, S Takamiya, K Suzuki, T Sato, H Oya, T Ozawa, Y Kagawa. Deficiencies in complex I subunits of the respiratory chain in Parkinson's disease. Biochem Biophys Res Commun 163:1450–1455, 1989.
- AHV Schapira, JM Cooper, D Dexter, P Jenner, JB Clark, CD Marsden. Mitochondrial complex I deficiency in Parkinson's disease. Lancet 1:1269, 1989.
- AHV Schapira, VM Mann, JM Cooper, D Dexter, SE Daniel, P Jenner, JB Clark, CD Marsden. Anatomic and disease specificity of NADH CoQ1reductase (complex I) deficiency in Parkinson's disease. J Neurochem 55:2142– 2145, 1990.
- 111. M Orth, AH Schapira. Mitochondrial involvement in Parkinson's disease. Neurochem Int 40:533–541, 2002.
- 112. R Betarbet, TB Sherer, G MacKenzie, M Garcia-Osuna, AV Panov, JT Greenamyre. Chronic systemic pesticide exposure reproduces features of Parkinson's disease. Nat Neurosci 3:1301–1306, 2000.
- Y Mizuno, K Suzuki, S Ohta. Postmortem changes in mitochondrial respiratory enzymes in brain and a preliminary observation in Parkinson's disease. J Neurol Sci 96:49–57, 1990.

- 114. MW Cleeter, JM Cooper, AH Schapira. Nitric oxide enhances MPP(+) inhibition of complex I. FEBS Lett 504:50–52, 2001.
- T Finkel, NJ Holbrook. Oxidants, oxidative stress and the biology of ageing. Nature 408:239–247, 2000.
- 116. Y Agid. Levodopa: is toxicity a myth? Neurology 50:858-863, 1998.
- 117. Y Agid. Levodopa. Is toxicity a myth? 1998. Neurology 57:S46-51, 2001.
- 118. J Jankovic. Levodopa strengths and weaknesses. Neurology 58:S19-32, 2002.
- D Berg, M Gerlach, MB Youdim, KL Double, L. Zecca, P Riederer, G Becker. Brain iron pathways and their relevance to Parkinson's disease. J Neurochem 79:225–236, 2001.
- M Gu, AD Owen, SE Toffa, JM Cooper, DT Dexter, P Jenner, CD Marsden, AH Schapira. Mitochondrial function, GSH and iron in neurodegeneration and Lewy body diseases. J Neurol Sci 158:24–29, 1998.
- MV Clement, LH Long, J Ramalingam, B Halliwell. The cytotoxicity of dopamine may be an artefact of cell culture. J Neurochem 81:414–421, 2002.
- 122. M Gerlach, D Blum-Degen, J Lan, P Riederer. Nitric oxide in the pathogenesis of Parkinson's disease. Adv Neurol 80:239–245, 1999.
- 123. GT Liberatore, V Jackson-Lewis, S Vukosavic, AS Mandir, M Vila, WG McAuliffe, VL Dawson, TM Dawson, S Przedborski. Inducible nitric oxide synthase stimulates dopaminergic neurodegeneration in the MPTP model of Parkinson disease. Nat Med 5:1403–1409, 1999.
- MJ LaVoie, TG Hastings. Peroxynitrite- and nitrite-induced oxidation of dopamine: implications for nitric oxide in dopaminergic cell loss. J Neurochem 73:2546–2554, 1999.
- AJ Nappi, E Vass. The effects of nitric oxide on the oxidations of l-dopa and dopamine mediated by tyrosinase and peroxidase. J Biol Chem 276:11214– 11222, 2001.
- 126. TL Perry, DV Godin, S Hansen. Parkinson's disease: a disorder due to nigral glutathione deficiency? Neurosci Lett 33:305–310, 1982.
- 127. SJ Kish, C Morito, O Hornykiewicz. Glutathione peroxidase activity in Parkinson's disease brain. Neurosci Lett 58:343–346, 1985.
- P Jenner. Presymptomatic detection of Parkinson's disease. J Neural Transm Suppl 40:23–36, 1993.
- 129. M Merad-Boudia, A Nicole, D Santiard-Baron, C Saille, I Ceballos-Picot. Mitochondrial impairment as an early event in the process of apoptosis induced by glutathione depletion in neuronal cells: relevance to Parkinson's disease. Biochem Pharmacol 56:645–655, 1998.
- N Jha, O Jurma, G Lalli, Y Liu, EH Pettus, JT Greenamyre, RM Liu, HJ Forman, JK Andersen. Glutathione depletion in PC12 results in selective inhibition of mitochondrial complex I activity. Implications for Parkinson's disease. J Biol Chem 275:26096–26101, 2000.
- 131. RH Swerdlow. Mitochondrial DNA-related mitochondrial dysfunction in neurodegenerative diseases. Arch Pathol Lab Med 126:271–280, 2002.
- 132. RH Swerdlow, JK Parks, JN Davis, 2nd, DS Cassarino, PA Trimmer, LJ Currie, J Dougherty, WS Bridges, JP Bennett, Jr., GF Wooten, WD Parker.

Matrilineal inheritance of complex I dysfunction in a multigenerational Parkinson's disease family. Ann Neurol 44:873–881, 1998.

- M Gu, JM Cooper, JW Taanman, AH Schapira. Mitochondrial DNA transmission of the mitochondrial defect in Parkinson's disease. Ann Neurol 44:177–186, 1998.
- 134. EM Grasbon-Frodl, S Kosel, M Sprinzl, U von Eitzen, P Mehraein, MB Graeber. Two novel point mutations of mitochondrial tRNA genes in histologically confirmed Parkinson disease. Neurogenetics 2:121–127, 1999.
- 135. RE Burke and NG Kholodilov. Programmed cell death: does it play a role in Parkinson's disease? Ann Neurol 44:S126–133, 1998.
- EC Hirsch, S Hunot, B Faucheux, Y Agid, Y Mizuno, H Mochizuki, WG Tatton, N Tatton, WC Olanow. Dopaminergic neurons degenerate by apoptosis in Parkinson's disease. Mov Disord 14:383–385, 1999.
- 137. D Blum, S Torch, N Lambeng, M Nissou, AL Benabid, R Sadoul, JM Verna. Molecular pathways involved in the neurotoxicity of 6-OHDA, dopamine and MPTP: contribution to the apoptotic theory in Parkinson's disease. Prog Neurobiol 65:135–172, 2001.
- DL Vaux, A Strasser. The molecular biology of apoptosis. Proc Natl Acad Sci USA 93:2239–2244, 1996.
- 139. SH Kaufmann, MO Hengartner. Programmed cell death: alive and well in the new millennium. Trends Cell Biol 11:526–534, 2001.
- NA Tatton. Increased caspase 3 and Bax immunoreactivity accompany nuclear GAPDH translocation and neuronal apoptosis in Parkinson's disease. Exp Neurol 166:29–43, 2000.
- 141. A Hartmann, PP Michel, JD Troadec, A Mouatt-Prigent, BA Faucheux, M Ruberg, Y Agid, EC Hirsch. Is Bax a mitochondrial mediator in apoptotic death of dopaminergic neurons in Parkinson's disease? J Neurochem 76:1785– 1793, 2001.
- 142. PA Trimmer, TS Smith, AB Jung, JP Bennett, Jr. Dopamine neurons from transgenic mice with a knockout of the p53 gene resist MPTP neurotoxicity. Neurodegeneration 5:233–239, 1996.
- 143. M Vila, V Jackson-Lewis, S Vukosavic, R Djaldetti, G Liberatore, D Offen, SJ Korsmeyer, S Przedborski. Bax ablation prevents dopaminergic neurodegeneration in the 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine mouse model of Parkinson's disease. Proc Natl Acad Sci USA 98:2837–2842, 2001.
- 144. D Offen, PM Beart, NS Cheung, CJ Pascoe, A Hochman, S Gorodin, E Melamed, R Bernard, O Bernard. Transgenic mice expressing human Bcl-2 in their neurons are resistant to 6-hydroxydopamine and 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine neurotoxicity. Proc Natl Acad Sci USA 95:5789– 5794, 1998.
- L Yang, RT Matthews, JB Schulz, T Klockgether, AW Liao, JC Martinou, JB Penney, Jr., BT Hyman, MF Beal. 1-Methyl-4-phenyl-1,2,3,6-tetrahydropyride neurotoxicity is attenuated in mice overexpressing Bcl-2. J Neurosci 18:8145–8152, 1998.

- B Cutillas, M Espejo, J Gil, I Ferrer, S Ambrosio. Caspase inhibition protects nigral neurons against 6-OHDA-induced retrograde degeneration. Neuroreport 10:2605–2608, 1999.
- 147. BS Jeon, NG Kholodilov, TF Oo, SY Kim, KJ Tomaselli, A Srinivasan, L Stefanis, RE Burke. Activation of caspase-3 in developmental models of programmed cell death in neurons of the substantia nigra. J Neurochem 73:322–333, 1999.
- 148. N Takai, H Nakanishi, K Tanabe, T Nishioku, T Sugiyama, M Fujiwara, K Yamamoto. Involvement of caspase-like proteinases in apoptosis of neuronal PC12 cells and primary cultured microglia induced by 6-hydroxydopamine. J Neurosci Res 54:214–222, 1998.
- RC Dodel, Y Du, KR Bales, Z Ling, PM Carvey, SM Paul. Caspase-3-like proteases and 6-hydroxydopamine induced neuronal cell death. Brain Res Mol Brain Res 64:141–148, 1999.
- H Turmel, A Hartmann, K Parain, A Douhou, A Srinivasan, Y Agid, EC Hirsch. Caspase-3 activation in 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine (MPTP)-treated mice. Mov Disord 16:185–189, 2001.
- 151. M Mogi, A Togari, T Kondo, Y Mizuno, O Komure, S Kuno, H Ichinose, T Nagatsu. Caspase activities and tumor necrosis factor receptor R1 (p55) level are elevated in the substantia nigra from parkinsonian brain. J Neural Transm 107:335–341, 2000.
- 152. A Hartmann, S Hunot, PP Michel, MP Muriel, S Vyas, BA Faucheux, A Mouatt-Prigent, H Turmel, A Srinivasan, M Ruberg, GI Evan, Y Agid, EC Hirsch. Caspase-3: a vulnerability factor and final effector in apoptotic death of dopaminergic neurons in Parkinson's disease. Proc Natl Acad Sci USA 97:2875–2880, 2000.
- 153. A Hartmann, JD Troadec, S Hunot, K Kikly, BA Faucheux, A Mouatt-Prigent, M Ruberg, Y Agid, EC Hirsch. Caspase-8 is an effector in apoptotic death of dopaminergic neurons in Parkinson's disease, but pathway inhibition results in neuronal necrosis. J Neurosci 21:2247–2255, 2001.
- 154. P Anglade, S Vyas, F Javoy-Agid, MT Herrero, PP Michel, J Marquez, A Mouatt-Prigent, M Ruberg, EC Hirsch, Y Agid. Apoptosis and autophagy in nigral neurons of patients with Parkinson's disease. Histol Histopathol 12:25– 31, 1997.
- 155. H Mochizuki, K Goto, H Mori, Y Mizuno. Histochemical detection of apoptosis in Parkinson's disease. J Neurol Sci 137:120–123, 1996.
- H Mochizuki, H Mori, Y Mizuno. Apoptosis in neurodegenerative disorders. J Neural Transm Suppl 50:125–140, 1997.
- 157. A Hartmann, A Mouatt-Prigent, BA Faucheux, Y Agid, EC Hirsch. FADD: a link between TNF family receptors and caspases in Parkinson's disease. Neurology 58:308–310, 2002.
- 158. SR Datta, A Brunet, ME Greenberg. Cellular survival: a play in three Akts. Genes Dev 13:2905–2927, 1999.